

RAPID SPECTROPHOTOMETRIC METHOD FOR HISTAMINE DETERMINATION IN FISH USING ALIZARIN RED S AND METAL

(Kaedah Spektrofotometri Pantas untuk Penentuan Histamin dalam Ikan Menggunakan Alizarin Merah S dan Logam)

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Abstract

An analytical method for the determination of histamine using metals and alizarin red S (ARS) reagents by UV-Visible spectrophotometry was developed. Cu(II), Co(II) and ARS were used to form colored complex with histamine. The developed method was used to detect qualitatively and quantitatively the presence of histamine. Absorbance of Cu(II) and Co(II) complex were measured at a maximum wavelength of 505.5 and 567 nm, respectively. Optimization of analytical parameters such as concentration of metals, concentration of ARS, sample solution pH, and optimum time for complex formation were performed. Experimental results showed that the optimum concentration of Cu(II) was 50 ppm, 75 ppm ARS, sample solution pH 6, and 15 minutes optimum time, while the optimum concentration of Co(II) was 125 ppm, ARS was 75 and 50 ppm, sample solution pH 5 and 10 minutes optimum time. Method validation indicated that the coefficient of variation, limit of detection (LOD) (S/N = 3), and limit of quantitation (LOQ = 10S/N) of Cu(II) were 0.65%, 8.94 ppm, and 29.82 ppm respectively with a sensitivity of 0.0054 ppm and linearity of 0.9959. Accuracy of histamine determination through Cu-ARS-histamine complex at a concentration of 50, 100, and 150 ppm were 105.7% (RSD = 0.777%, n = 3), 105.06% (RSD = 0.606%, n = 3), and 94.12% (RSD = 0.767%, n = 3), respectively. Meanwhile, the CV, LOD, and LOQ of Co(II) were 0.28%, 2.58 ppm; and 8.6 ppm respectively with a sensitivity of 0.0006 ppm and linearity of 0.9965. Accuracy of histamine determination through Co-ARS-histamine complex at a concentration of 50, 75, and 125 ppm were 98.33%, 95.83%, 93.88%, respectively. The proposed method was successfully applied to the determination of histamine in a fish sample with quantitative recovery; for Cu(II) complex (99.23%) and Co(II) complex (102.62%).

Keywords: histamine, metals, Cu(II), Co(II), ARS, UV-visible spectrophotometry

Abstrak

Kaedah analisis untuk penentuan histamin menggunakan logam dan reagen alizarin merah S (ARS) menggunakan spektrofotometri ultralembayung-nampak telah dibangunkan. Cu(II), Co(II), dan ARS digunakan untuk membentuk kompleks berwarna dengan histamin. Kaedah yang telah dibangunkan digunakan untuk mengesan histamin secara kualitatif dan kuantitatif. Serapan kompleks Cu(II) dan Co(II) diukur pada panjang gelombang maksimum masing-masing 505.5 dan 567 nm. Pengoptimuman parameter analisis seperti kepekatan logam, kepekatan ARS, pH larutan sampel, dan masa optimum bagi pembentukan kompleks telah dilakukan. Keputusan eksperimen menunjukkan bahawa kepekatan optimum Cu(II) ialah 50 ppm, 75 ppm ARS, pH 6 larutan sampel, dan masa optimum 15 minit, manakala kepekatan optimum Co(II) ialah 125 ppm, ARS ialah 75 dan 50 ppm, pH larutan sampel 5 dan 10 minit masa optimum. Pengesahan kaedah menunjukkan bahawa pekali variasi (CV),

had pengesanan (LOD) ($S/N = 3$) dan had kuantitatif (LOQ = 10 S/N) Cu(II) masing-masing adalah 0.65%, 8.94 ppm dan 29.82 ppm dengan sensitiviti 0.0054 ppm dan kelinearan 0.9959. Ketepatan penentuan histamin melalui kompleks Cu-ARS-histamin pada kepekatan 50, 100, dan 150 ppm ialah 105.7% (RSD = 0.777%, $n = 3$), 105.06% (RSD = 0.606%, $n = 3$), dan 94.12% (RSD = 0.767%, $n = 3$), sementara itu, CV, LOD, dan LOQ Co(II) masing-masing adalah 0.28%, 2.58 ppm dan 8.6 ppm dengan kepekaan 0.0006 ppm dan kelinearan 0.9965. Ketepatan penentuan histamin melalui kompleks Co-ARS-histamin pada kepekatan 50, 75, dan 125 ppm masing-masing adalah 98.33%, 95.83%, dan 93.88%. Kaedah yang dicadangkan berjaya digunakan untuk menentukan histamin dalam sampel ikan dengan pengembalian kuantitatif untuk kompleks Cu(II) (99.23%) dan kompleks Co(II) (102.62%).

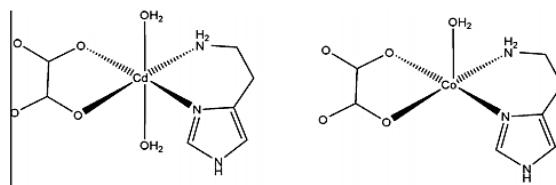
Kata kunci: histamin, logam, Cu(II), Co(II), alizarin merah S, spektrofotometri ultralembayung-nampak

Introduction

Histamine with chemical formula $C_5H_9N_3$ is a small-molecule hormone present in essentially all mammalian tissues which acts at the central nervous system level and in the regulation of sleep [1]. Histamine is produced in species of the *Scombroideae* family such as tuna and mackerel, mahi-mahi, blue fish, and sardine [2]. Histamine is one of the most toxic and most commonly found in foods and its concentration is frequently regarded as one of the biomarkers for quality control during the food production, transportation and marketing [3, 4]. Nowadays, an increased consumption of fish has been recommended due to its health promoting properties. However, some fish can cause histamine poisoning if its quality is not assured [5].

As a result, there were some regulations about histamine level in food. Based on Standard Nasional Indonesia (SNI) 01-2360 year 2008, histamine maximum limit in Indonesian fish is 100 mg kg^{-1} . The European Commission has made a regulation of safety requirements on the histamine presence in seafood with amounts lower than 10 mg/kg of histamine which indicate that fish has good quality, a level of 30 mg/kg suggests significant deterioration, and a content of 50 mg/kg or higher is an evidence of definite decomposition [6]. While the United States Food and Drug Administration (FDA) has established 50 ppm of histamine as the chemical index for fish spoilage [7]. Based on the assessment of poisoning cases, the guidance levels suggested for histamine content in seafood are: $<50 \text{ ppm}$ for safe consumption; $200\text{--}1000 \text{ ppm}$ as probably toxic; and $>1000 \text{ ppm}$ as toxic and unsafe for human consumption [8, 9].

Several analytical methods have been developed for the analysis and determination of histamine levels such as ultrasensitive flow injection electrochemical [10], lignin modified glassy carbon electrode [11], thin layer chromatography (TLC) [12], high performance liquid chromatography (HPLC) [13], ^1H NMR [14], electrochemiluminescence [15], liquid chromatography tandem mass spectrometry [16], polymerase chain reaction/denaturing gradient gel electrophoresis (PCR-DGGE) [3] and spectrophotometry [17]. Complex reaction between histamine, Co(II) and oxalate has been reported [18] with the complex structure shown (Figure 1), but it has not given a significant change of wavelength and solution color. On the other hand, alizarin red S (ARS) with chemical structure shown (Figure 2) is well known to form colored complex with metallic and organic compounds such as Al(III) [19], Cu [20], Co and Ni [21], Cu, Co, Ni and cysteine [22], and Ni-histamine [23] with a wide range of ARS solvents.



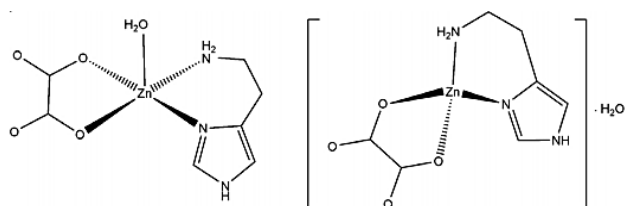


Figure 1. Complex structure of histamine-metal-oxalate [19]

The presence of a quinoid oxygen, with two hydroxyl groups at the α - and β -positions in ARS makes it very suitable for chelation, and its derivatives are well known colorimetric reagents with several applications. Furthermore, the presence of sulphonate groups in the molecule improves its reactivity [25]. ARS dye can exist in different forms depending on the type of media used, so this study used ARS as a complexing and coloring agent in the determination of histamine.

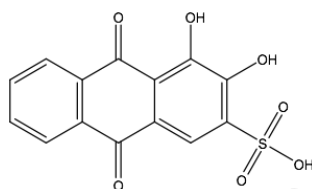


Figure 2. Chemical structure of ARS

In this study, a novel spectrophotometric method for determination of histamine has been developed based on histamine-metal-ARS complex formation. Analytical procedure for the determination of histamine is based on the complex's reaction, which can form a colored compound and then the absorbance was measured using UV-visible spectrophotometer. Optimization of metal concentration, ARS concentration, solution pH and time for formation of complexes have been investigated as well. The analytical validation parameters namely LOD, LOQ, sensitivity, coefficient of variation (CV), linearity, and accuracy were performed. The validated method was then applied to the determination of histamine in fish sample.

Materials and Methods

Materials

Histamine reference standard was purchased from Sigma Aldrich (Science Park Drive, Singapore). Copper sulphate hexahydrate, cobalt(II) nitrate hexahydrate, and ARS were purchased from Merck (Darmstadt, Germany). Methanol, sodium acetate, sodium hydrogen phosphate, sodium dihydrogen phosphate, acetic acid, trichloroacetic acid, and sodium hydroxide were pure analytical grade. Whatman filter paper 42 from Merck (Darmstadt, Germany) was used to filtering the sample extract.

Instrumentation

A Shimadzu-1800 UV-Visible spectrophotometer (Kyoto, Japan) was used to measure the wavelength and absorbance of the complexes formed. ThermoFisher Scientific Eutech 700 pH meter from (Massachusetts, USA) was used to measure the pH of the buffer solution prepared and standard solution for analytical optimization of solution pH.

Preparation of reagent

Histamine (0.1000 g) was weighed and dissolved in methanol, then transferred to a 100 mL volumetric flask and made up to the mark with the same solvent. $\text{CuSO}_4 \cdot 6\text{H}_2\text{O}$ (0.3929 g) was weighed and dissolved in distilled water. The solution was transferred to a 100 mL volumetric flask and made up to mark with distilled water. $\text{Co}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ (0.4940 g) was weighed and dissolved in distilled water. The solution was transferred to a 100 mL volumetric flask and made up to mark with distilled water. All solutions were shaken to homogenize. The same procedure was performed for ARS with a weight of 0.1 gram. Buffer solutions used to make an acidic and alkaline

medium were phosphate buffer solution and acetic buffer solution. Phosphate buffer solution for pH 6, 7, and 8 was made from 1 M sodium hydrogenphosphate and sodium dihydrogen phosphate solution. While acetic buffer solution for pH 4 and 5 was made from 1 M acetic acid and trichloroacetic acid solution.

Preparation of ARS-metal complex solution

A 0.25 mL ARS stock solution 1000 ppm and 1.0 mL pH 7 buffer solution were transferred quantitatively into a 10 mL volumetric flask and added with 1.0 mL Cu(II) stock solution 1000 ppm, respectively. It was made up to the mark with methanol, and shaken until homogeneous and left for 10 min. The wavelength of complex solution formed was scanned from 300-800 nm and maximum absorbance was noted. The same procedure was repeated for Co(II) solution.

Preparation of histamine-ARS-metal complex solution

A 0.25 mL ARS stock solution 1000 ppm, 1.25 mL of histamine stock solution 1000 ppm, 1 mL buffer solution of pH 7 buffer solution were transferred into a 10 mL volumetric flask and added with 1.0 mL Co(II) stock solution 1000 ppm, left at room temperature for 10 min. It was made up to mark with methanol. The wavelength of complex solution formed was scanned from 300 - 800 nm and maximum absorbance was noted. The same procedure was repeated for Cu(II) solution.

Optimization of metal concentration

4 sets of 10 mL volumetric flask were prepared, and each volumetric flask was added with 1.25 mL histamine stock solution. In each volumetric flask, then added with Cu(II) stock solution with variation volume of 0.25; 0.50; 0.75; and 1.00 mL 1000 ppm, hence the concentration of metal was 25, 50, 75, and 100 ppm; then added with 1.00 mL pH 7 buffer solution; and 0.25 mL ARS stock solution 1000 ppm. Another 4 pieces of 10 mL volumetric flask was added with 1.25 mL histamine stock solution; then added with 1.00 mL buffer solution pH 7; 0.25 mL alizarin red S and 1.00; 1.25 and 1.50 mL of Co(II) stock solution. Then in each volumetric flask, the solution was diluted with methanol to mark and homogenized. The solution was allowed to stand for 10 minutes and its absorbance was measured at the maximum wavelength. A plot of the relationship between the concentration of alizarin red S and absorbance was made. The optimum concentration of the metal was obtained from the concentration value of the solution having the greatest absorbance value.

Optimization of ARS concentration

8 sets of 10 mL volumetric flask were prepared, and each volumetric flask was added with 1.25 mL histamine stock solution. In each volumetric flask was added with metal aqueous solution according to the optimum concentration and then added with 1.00 mL buffer solution pH 7; and 0.25, 0.50, 0.75, and 1.00 mL of ARS stock solution 1000 ppm, hence the concentration of ARS was 25, 50, 50, 75, and 100 ppm. The solution was diluted with methanol to the mark and homogenized. The solution was allowed to stand for 10 minutes and its absorbance was measured at the maximum wavelength. A curve of the relationship between the concentration of red alizarin S and absorbance was made. The optimum concentration of the metal was obtained from the concentration value of the solution having the greatest absorbance value.

Optimization of pH solution

10 sets of 10 mL volumetric flask were prepared. Each volumetric flask was added 1.25 mL of histamine stock solution and metal aqueous solution according to the optimum concentration, ARS with optimum concentration, and then added with 1.00 mL buffer pH solution of 4, 5, 6, 7, and 8 in each flask. The solution was diluted with methanol to the mark and homogenized. The solution was allowed to stand for 10 minutes and its absorbance measured at the maximum wavelength. A curve of the relationship between the concentration of red alizarin S and absorbance was made. The optimum pH was obtained from the pH value of the solution having the greatest absorbance value.

Optimization of time

10 sets of 10 mL volumetric flask were prepared, and each volumetric flask was added 1.25 mL of histamine stock solution. In each volumetric flask was added a solution of metal in accordance with the optimum concentration, ARS with optimum concentration, and then added with 1.00 mL of optimum buffer pH solution. The solution was

allowed to stand for 10 minutes and its absorbance measured at the maximum wavelength. A curve of the relationship between the concentration of red alizarin S and absorbance was made. The optimum time was obtained from the time value of the solution having the greatest absorbance value.

Determination of histamine in fish

A sample preparation as follows was tried to detect and determine the histamine in fish fillets according to the previous method [25]. Briefly, 10 g fish muscle was minced with 10 mL of 10% trichloroacetic acid in water using a homogenizer. The homogenate was spiked with histamine standard, resulting in artificial contamination at 50, 100, and 150 mg/kg. The homogenate neutralized by 0.1 mL 0.1 M NaOH solution was centrifuged at 3000 r/min for 5 minutes. Then 2 mL of the supernatant was then filtered through a Whatman filter paper 42. The supernatant then used to determination of histamine in fish sample with the optimum condition for each metal.

Results and Discussion

A spectrophotometric method for the determination of histamine is based on the complex reaction between histamine, ARS, and metal using an UV-visible spectrophotometer with a concentration of histamine 125 ppm, ARS 25 ppm, and metal(II) 100 ppm has been done. From Figure 3, it can be seen that the complex compounds of ARS and Cu(II) and Co(II) produce maximum wavelength at 544 nm and 543 nm, respectively. Structure of ARS-metal in various media which has been found before by the simple Hückel method [26] indicates that alizarin complexation occurs at the peri-hydroxy carbonyl group and results in the formation of the $C=O \rightarrow M-O$ coordination bond as shown in Figure 4(a). In acidic media, O-M bond remains covalent and the electronic absorption spectra of alizarinates are identical to the spectrum of nonionized alizarin as shown in Figure 4(b) in which H atom is replaced with metal atom. In alkali solution, alizarin reacted with metal ions as the *ortho*-diphenol compound as shown in Figure 4(c).

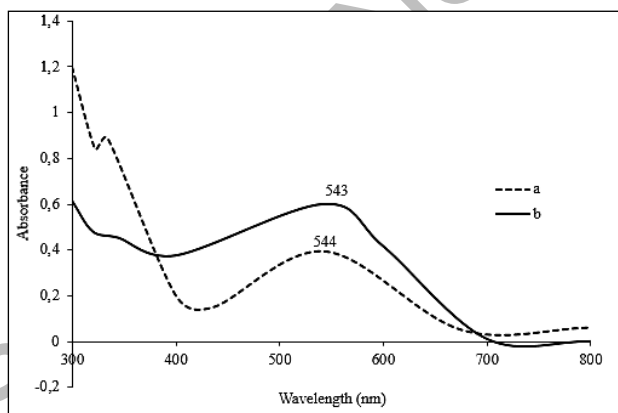


Figure 3. (a) UV-visible spectra of ARS-Cu(II) complex (b) ARS-Co(II) complex

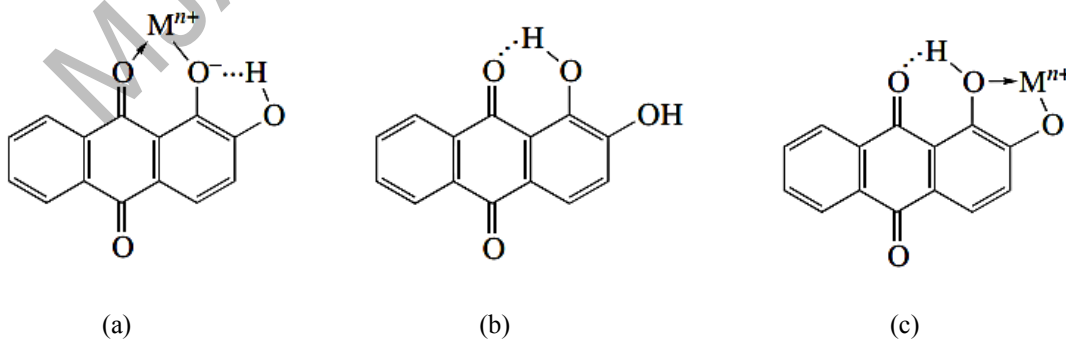


Figure 4. Structure of ARS complex in various medium (a) neutral medium (b) acidic medium (c) alkaline medium [26]

This complex compound will then react with histamine forming a new pink complex compound with a maximum wavelength for Cu(II) and Co(II) at 505.5 nm; and 567 nm, respectively. These results indicate that a shift in the wavelength of maximum absorption into lower wavelength due to hypsochromic effect for Cu(II) and greater due to bathochromic effect for Co(II) and electron configuration of Co and Cu as shown in Figure 5. The metals bind the existing amine group on histamine and carbonyl group of ARS. This compound is predicted that it has an octahedral shaped molecular. Based on crystal field theory, octahedral shape has an energy splitting as shown in Figure 6 when stabilizing energy of metal-ligand complex due to the amount of electrons in t_{2g} meanwhile unstabilizing energy of metal-ligand complex is due to the amount of electrons e.g. Cu and Co have the same amount of electrons in t_{2g} but it has different amount of electrons e.g. Cu has three electrons while Co has an electron there. It means that Cu is more unstable than Co, so Cu needed more energy to form the complex than Co. Furthermore, it would give a greater wavelength. In the molecular structure of the complex as shown in Figure 7, histamine acts as a bidentate ligand by chelating the metal ion. The metal is in a square pyramidal geometry defined by two N atoms of aqua ligand at apex of pyramid. Histamine along with neighboring carbonyl ion participates in weak hydrogen bonds of N–H–O in the crystal packing of $[\text{Metal}(\text{His})(\text{ARS})(\text{H}_2\text{O})_2]$ complex.

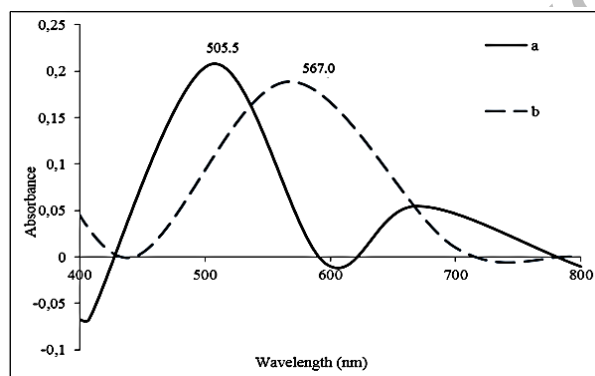


Figure 5. (a) UV-visible spectra of ARS-histamine-Cu(II) complex (b) ARS-histamine-Co(II) complex

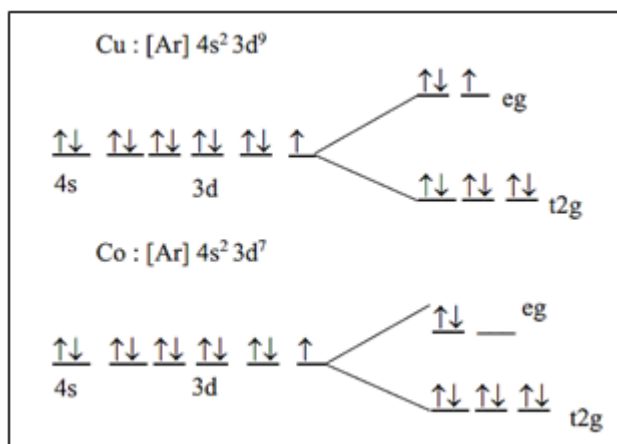


Figure 6. Splitting energy of Cu and Co due to crystal field theory

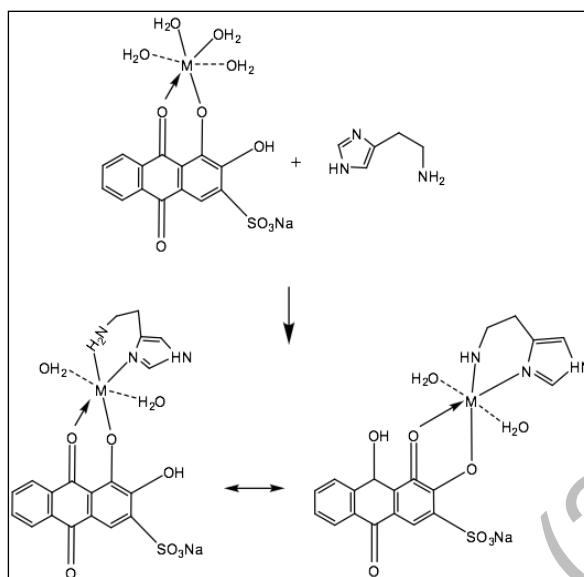
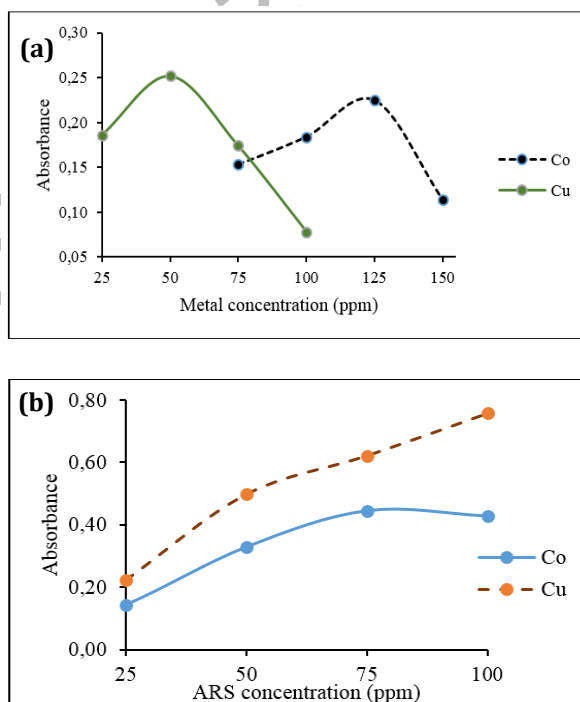


Figure 7. The prediction reaction of metal(II), histamine and ARS

The wavelength of the histamine - ARS - metal complex compound is used as the wavelength for the entire analytical parameter optimization. Optimization of metal and ARS concentration is obtained from the increase in absorbance due to higher concentration. The optimum results of Cu(II) and Co(II) concentration are 50 and 125 ppm as shown in Figure 8(a) and (b), respectively. Optimization of ARS for Cu(II) and Co(II) complex is 75 ppm as shown in Figure 8(b).



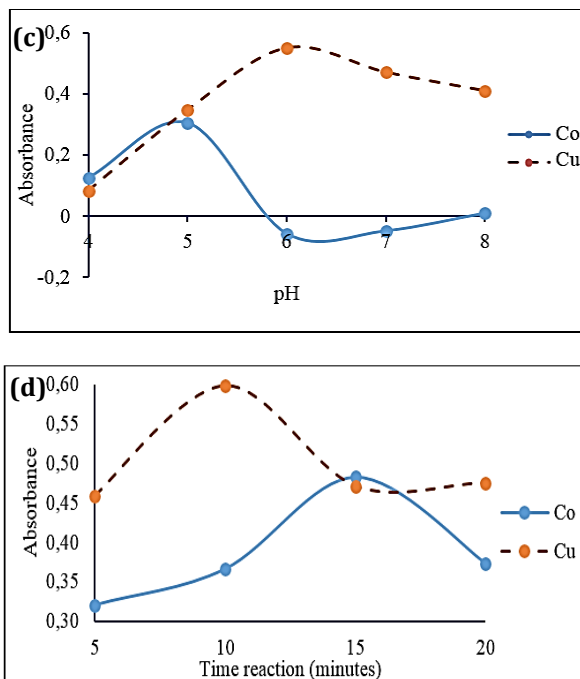


Figure 8. Optimization of (a) metal concentration (b) ARS concentration (c) pH solution (d) time due to the higher absorbance

The optimum pH for complex formation was determined. In Figure 8(c), it is obtained that pH optimum is at pH 6 and 5 for Cu(II) and Co(II), respectively. The reaction of Cu(II) and Co(II) complex formation took place in acidic conditions because in $\text{pH} > 7$ histamine - ARS - Cu(II) and $\text{pH} > 6$ Co(II) solution lead the precipitation of $\text{M}(\text{OH})_2$ and starts to form colloidal solution. It is because in this condition, $K_{sp} \text{M}(\text{OH})_2$ has been reached [27]. Time of Cu(II) and Co(II) complex are 10 and 15 minutes, respectively, as shown in Figure 8(d).

From the results of the optimization (concentration of ARS and metal(II), pH and time), linear calibration curves are obtained at the range of 25 - 150 ppm of histamine as shown in Figure 9. The linear regression for Cu(II) and Co(II) is $y = 0.0054x - 0.1944$ and $R^2 = 0.9959$; $y = 0.0006x + 0.3136$ and $R^2 = 0.9965$, respectively. Validation parameters for each metal are shown in Table 1. Based on linear regression and Table 1, linearity is closed to 1 so it is quite good. Two directions ANOVA shows $t_{\text{table}} < t_{\text{calculation}}$, so H_0 is rejected and there is a correlation between the concentration with absorbance. The accuracy of the method range of 101.63% and 96.01% for Cu(II) and Co(II) complex is shown in the Table 2 so it is indicated that this method of approaching the actual state of histamine concentration. Variation coefficient of histamine by 0.650% and 0.275% is shown in Table 3 for each metal. Furthermore, Cu(II) complex has higher sensitivity than Co(II) with a sensitivity of 0.0054; while sensitivity of Co(II) complex is 0.0006 obtained from the slope of histamine standard curve.

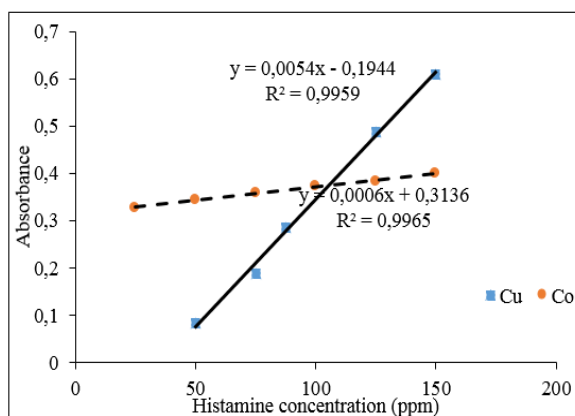


Figure 9. Calibration curve of histamine

Table 1. Validation parameters of method

Validation Parameter	Result	
Metal	Cu(II)	Co(II)
LOD	8.94 ppm	2.58 ppm
LOQ	29.82 ppm	8.6 ppm
Linearity	0.99	0.99
Sensitivity	0.0054/ppm (slope)	0.0006/ppm (slope)

Table 2. Accuracy of method for Cu(II) and Co(II) complex

Actual Concentration (ppm)	Cu(II)		Co(II)	
	Found Concentration (ppm)	Recovery (%)	Found Concentration (ppm)	Recovery (%)
50	52.935	105.70	49.167	98.33
100	105.060	105.06	95.830	95.83
150	121.513	91.12	140.830	93.88
	Mean	101.63	Mean	96.01

Table 3. Variation coefficient (precision) of histamine standard for Cu(II) and Co(II) complex

Histamine (ppm)	Cu(II)				Precision (%)	Co(II)				Precision (%)
	Absorbance					Absorbance				
	x ₁	x ₂	x ₃			x ₁	x ₂	x ₃		
50	0.090	0.091	0.091	0.091	0.777	0.344	0.343	0.342	0.343	0.291
100	0.303	0.303	0.301	0.303	0.406	0.371	0.373	0.369	0.371	0.539
150	0.566	0.565	0.573	0.568	0.767	0.398	0.398	0.398	0.398	0.000
	Mean				0.650	Mean				0.275

The optimum result of method has been used for histamine determination in fish. The sample which known previously of histamine levels was spiked using histamine standard solutions at concentrations of 50, 100, and 150 mg/kg. Then the mixed solution was determined using the developed spectrophotometric method as shown in Table 4. Recovery results obtained was close to 100%, it means that both of these metals complexes have a good recovery to be applied to fish samples.

Table 4. Sample application of Cu(II) and Co(II) complex

Real Concentration (ppm)	Cu(II)		Co(II)	
	Found Concentration (ppm)	Recovery (%) (n = 3)	Found Concentration (ppm)	Recovery (%) (n = 3)
Fish	52	-	52	-
Fish + 50	100	97.72	105	102.71
Fish + 100	149	98.26	157	103.03
Fish + 150	202	101.71	206	102.12
	Mean	99.23	Mean	102.62

Conclusion

The spectrophotometric method for the determination of histamine based on complex reaction using metals and ARS as reagent has been developed which has simple methodology, easy work-up, applicable, and accurate.

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References

1. Kaştaş, G., Paşaoğlu, H., Karabulut, B. and Bulut, İ. (2010). Structural and spectroscopic studies on cadmium complex of a biogenic amine, histamine. *Chemical Physics Letters*, 488 (4-6): 162–167.
2. Bulushi, I. A., Poole, S., Deeth, H. C. and Dykes, G. A. (2009). Biogenic amines in fish: Roles in intoxication, spoilage, and nitrosamine formation—a review. *Critical Reviews in Food Science and Nutrition*, 49(4): 369–377.
3. Diaz, M., Ladero, V., Redruello, B., Sanchez-Llana, E., Rio, B., Fernandez, M., Martin, M. C. and Alvarez, M. A. (2016). A PCR-DGGE method for the identification of histamine-producing bacteria in cheese. *Food Control*, 63: 216-223.
4. Altieri, I., Semeraro, A., Scalise, F., Calderari, I. and Stacchini, P. (2016). European official control of food: Determination of histamine in fish products by a HPLC–UV–DAD Method. *Food Chemistry*. 211: 694–699.
5. Evangelista, W. P., Silva, T. M., Guidi, L. R., Tette, P. A. S., Byrro, R. M. D., Silva, P. S., Fernandes, C. and Gloria, M. B. A. (2016). Quality assurance of histamine analysis in fresh and canned fish. *Food Chemistry*, 211: 100-106.
6. Veseli, A., Vasjari, M., Arbnesi, T., Hajrizi, A., Svorc, L., Samphao, A. and Kalcher, K. (2016). Electrochemical determination of histamine in fish sauce using heterogeneous carbon electrodes modified with rhenium(IV) oxide. *Sensors and Actuators B*. 228: 774–781.
7. Department of Health and Human Services, Public Health Service, Food and Drug Administration, Center for Food Safety and Nutrition (2011). Fish and fishery products hazards and controls guidance, 4th edition 2011, April.
8. Lehane, L. and Olley, J. (2000). Review histamine fish poisoning revisited. *International Journal of Food Microbiology*, 58 (1-2): 1–37.

9. Keow, C. M., Bakar, F. A., Salleh, A. B., Heng, L. Y., Wagiran, R., Bean, L. S. (2007). An amperometric biosensor for the rapid assessment of histamine level in tiger prawn (*Penaeus monodon*) spoilage, *Food Chemistry*, 105(4): 1636–1641.
10. Akbari-adergani, B., Norouzi, P., Ganjali, M. R., and Dinarvand, R. (2010). Ultrasensitive flow-injection electrochemical method for determination of histamine in tuna fish sample. *Food Research International Journal*, 43(4): 1116–1122.
11. Degefu, H., Amare, M., Tessema, M. and Admassie, S. (2014), Lignin modified glassy carbon electrode for the electrochemical determination of histamine in human urine and wine samples. *Electrochimica Acta Journal*, 121: 307–314.
12. Tao, Z., Sato, M., Han, Y., Tan, Z., Yamaguchi, T. and Nakano, T. (2011). A simply and rapid method for histamine analysis in fish and fishery products by TLC determination. *Food Control*, 22(8): 1154–1577.
13. Wang, Z., Wu, J., Wu, S. and Bao, A. (2013). High performance liquid chromatographic determination of histamine in biological samples: the cerebrospinal fluid challenge—a review. *Analytica Chimica Acta*, 24(774): 1–10.
14. Henry, B., Gizzi, P. and Delpuech, J. (2015), Magnetic Non-equivalence and dynamic NMR of N-methylene protons in a histamine-containing pseudopeptide: Alanyl-glycyl-histamine. *Tetrahedron*, 71(36): 6227–6244.
15. An, D., Chen, Z., Zheng, J., Chen, S., Wang, L. and Su, W. (2016). Polyoxometalate functionalized tris(2,2-bipyridyl)dichlororuthenium(II) as the probe for electrochemiluminescence sensing of histamine, *Food Chemistry*, 194: 966–971.
16. Chimalakonda, K. C., Pang, E., Weaver, J. L., Howard, K. E., Patel, V. and Boyne II, M. T. (2015). Development and validation of a liquid-chromatography tandem mass spectrometry method to determine in vitro and in vivo histamine release, *Journal of Pharmaceutical and Biomedical Analysis*, 102: 494–499.
17. Jancsó, A., Selmeczi, K., Gizzi, P., Nagyc, N. V., Gajda, T. and Henry, B. (2011). The role of terminal amino group and histidine at the fourth position in the metal ion binding of oligopeptides revisited copper(II) and nickel(II) complexes of glycyl-glycyl-glycyl-histamine and its n-boc protected derivative. *Journal of Inorganic Biochemistry*, 105(1): 92–101.
18. Kaştaş, G., Pasaoglu, H. and Karabulut, B. (2011). Magnetic, structural and computational studies on transition metal complexes of a neurotransmitter, histamine. *Journal of Molecular Structure*, 1000(1-3): 39–48.
19. Sathish, R. S., Kumar, M. R., Rao, G. N., Kumar, K. A. and Janardhana, C. (2007). A water-soluble fluorescent fluoride ion probe based on ARS–Al(III) complex. *Spectrochimica Acta Part A*, 66: 457–461.
20. Panahi, H. A., Karimi, M., Moniri, E. and Soudi, H. (2008). Development of a sensitive spectrophotometric method for determination of copper. *African Journal of Pure and Applied Chemistry*, 2(10): 96–99.
21. Rohilla, R. and Gupta, U. (2012) Simultaneous determination of cobalt(ii) and nickel(ii) by first order derivative spectrophotometry in micellar media. *E-Journal of Chemistry*, 9(3): 1357–1363.
22. Gahlan, A. A., El-Mottaleb, M.A., Badawy, N. A., Kamale, F. H. and Ali, S. H. (2014). spectrophotometric studies on binary and ternary complexes of some metal ions with ARS and cysteine. *International Journal of Advanced Research*, 2(10): 570–584.
23. Jannatin, M., Supriyanto, G., Pudjiastuti, P. (2017). A novel spectrophotometric method for the determination of histamine based on its complex reaction with Ni(II) and ARS. *Indonesian Journal of Chemistry*, 17(1): 139–143.
24. Ali, N. M., Amaniampong, P. N. and Karam, A. (2016). Determination of optimal conditions for electrodeposition of tin(II) in the presence of ARS. *Heliyon*, 2(12): 1–26.
25. Xie, Z., Wang, Y., Chen, Y., Xu, X., Jin, Z. and Ding, Y. (2017). Tuneable surface enhanced raman spectroscopy hyphenated to chemically derivatized thin-layer chromatography plates for screening histamine in fish. *Food Chemistry*, 230: 547–552.
26. Fain, V. Y., Zaitsev, B. E. and Ryabov, M. A. (2004). Metal complexes with alizarin and ARS: Electronic absorption spectra and structure of ligands. *Russian Journal of Coordination Chemistry*, 30(5): 365–370.
27. Day, R. A. and Underwood, A. L. (2002). Analisis kimia kuantitatif. Edisi keenam. Penerbit Erlangga, Jakarta, Indonesia.