MALAYSIAN JOURNAL OF ANALYTICAL SCIENCES

Published by The Malaysian Analytical Sciences Society

ISSN 1394 - 2506

A-NOR STEROIDS FROM THE MARINE SPONGE, Clathria SPECIES

(Steroid A-nor daripada Span Laut, Spesies *Clathria*)

Idin Sahidin¹*, Carla Wulandari Sabandar¹, Wahyuni¹, Rini Hamsidi¹, Muhammad Hajrul Malaka¹, Baru Sadarun², La Ode Aslan²

¹Faculty of Pharmacy
²Faculty of Fisheries and Marine Science
Universitas Halu Oleo, Kendari 93232, Indonesia

*Corresponding author: sahidin02@yahoo.com

Received: 22 August 2017; Accepted: 20 March 2018

Abstract

Chemical compounds classified as A-nor steroids (1–4) were isolated from a marine sponge of the *Clathria* sp., collected from Southeast Sulawesi, Indonesia. Of these, one new compound, clathruhoate (1) was elucidated as 3β -(butyryloxymethyl)-A-nor- 5α -cholestane. All compounds were obtained using silica gel chromatography techniques and their structures were determined based on 1D and 2D NMR spectroscopic measurements and comparison with values from literature. Meanwhile, all compounds were inactive for antimicrobial activity.

Keywords: Clathria sp., marine sponge, A-nor steroid, 3β -(Butyryloxymethyl)-A-nor- 5α -cholestane

Abstrak

Sebatian kimia diklasifikasikan sebagai steroid A-nor (1–4) telah diasingkan daripada span laut Clathria yang dikumpul dari Sulawesi Tenggara, Indonesia. Satu sebatian baharu, klathruhoat (1) telah ditentukan sebagai 3β -(butiriloksimetil)-A-nor- 5α -kolestan. Semua sebatian diperoleh dengan menggunakan teknik kromatografi gel silika dan struktur sebatian-sebatian ini telah ditentukan menggunakan spektroskopi 1D dan 2D NMR dan dengan perbandingan nilai dari kajian literatur. Sementara itu, semua sebatian didapati tidak aktif terhadap aktiviti antimikrob.

Kata kunci: Clahtria sp., span laut, steroid A-nor, 3β -(Butiriloksimetil)-A-nor- 5α -kolestan

Introduction

The diversity of marine species in the Indonesian Archipelago has been recognized worldwide [1, 2], and various species of sponges have been discovered in east Indonesia [3, 4]. The genus *Clathria* from the family Microcionidae is classified as Demospongia and contains approximately 544 accepted species worldwide [5]. Of these, some species from the marine regions of Australia, Argentina, Indo-Pacific, Korea, Panama, Philippines and New Zealand have been phytochemically investigated and found to contain alkaloids [6–12], amides [13], peptides [14, 15], anthraquinones [16], and terpenoids [17–23]. Meanwhile, biological activities including antibacterial [7, 11, 21, 22], anti-HIV [18], cytotoxicity towards cancer cell lines [9, 10, 13, 14, 22], anti-inflammatory [19], and antiplasmodial [24] have been attributed to the species in the genus *Clathria*.

In our extensive study on the diversity and chemistry of Indonesian marine sponges, we investigated the chemical constituents of *Clathria* sp. from the Southeast Sulawesi marine region. The investigation resulted in the isolation and elucidation of one new *A*-nor steroid (1), together with three known *A*-nor steroids (2–4). The isolated compounds (1–4) were also evaluated for their antimicrobial activity.

Materials and Methods

Chemicals

Solvents used were distilled technical grades. Aluminium sheets kieselgel 60 PF $_{254}$ 0.25 mm (Merck 1.05554), silica gel 60 HF $_{254}$ 5-40 μ m (Merck 1.07747), and silica gel 60 GF $_{254}$ 5-40 μ m containing gypsum (Merck 1.07749) were used to perform thin layer chromatography (TLC), vacuum liquid chromatography (VLC), and radial chromatography (RC), respectively. Nutrient agar (NA), nutrient broth (NB), potato dextrose agar (PDA), potato dextrose broth (PDB), chloramphenicol, ketoconazole, and DMSO were used for antimicrobial assays.

Animal material

Samples of *Clathria* sp. (Figure 1) were collected by hand using SCUBA (Self Contained Underwater Breathing Apparatus) from the reef slope area (70°) of the Bintang Samudra Marine Education Park, Southeast Sulawesi, Indonesia, at a depth of 2–10 m, in May 2015. Samples (6.5 kg) were stored in ice and brought back shortly afterwards to the laboratory for further analysis. A sponge specimen was identified by the staffs of the Faculty of Fisheries and Marine Science of Universitas Halu Oleo with the registration number UHO-2015-03.

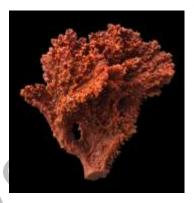


Figure 1. A photograph of the investigated *Clathria* sp.

Extraction and isolation

The dried powdered sponge (4 kg) was macerated with MeOH (3 x 8 L, 24 h each time) at room temperature and filtered. The solvent was evaporated under reduced pressure to yield the MeOH extract in the form of a brownish-yellow gum (300 g). A portion of this extract (30 g) was fractionated using silica gel vacuum liquid chromatography (VLC) (13 x 5 cm) with a gradient elution of *n*-hexane/EtOAc (9:1, 8:2, 7:3, 5:5, v/v) to produce five main fractions (F1-F5). Fraction F1 (2 g) was subjected to a silica gel radial chromatography (RC) and eluted isocratically with *n*-hexane/EtOAc (9.5:0.5, v/v) to yield nine subfractions (F1₁-F1₉). The combined subfractions F1₁-F1₃ (1.36 g) were further separated on a silica gel RC with an elution of *n*-hexane/CHCl₃ (9.5:0.5, v/v) to afford compound **1** (30 mg). Purification of subfraction F1₄ (0.14 g) using a silica gel RC with *n*-hexane/CH₂Cl₂ (6:4, v/v) as its mobile phase yielded compound **2** (23 mg). Fraction F2 (10 g) was subjected to a silica gel VLC (10 x 5 cm), eluted with *n*-hexane/EtOAc (9:1-0:10, v/v), and given eight subfractions (F2₁-F2₈). Subfraction F2₄ (0.8 g) was purified using silica gel RC with *n*-hexane/EtOAc/acetone (7:1:0.5, v/v) as its mobile phase to give compound **3** (200 mg). Purification of subfraction F2₆ (1.1 g) on a silica gel RC with the elution of *n*-hexane/EtOAc/acetone (7:1:0.5, v/v) yielded compound **4** (120 mg).

Characterization study

The infrared (IR) analysis of compounds was performed using a Nicolet iS5 spectrophotometer with iD5 ATR (Thermo Scientific, Waltham, MA, USA). The HRESI-MS spectra were measured using a Waters QTOF mass spectrometer (Milford, MA, USA). Meanwhile, the ¹H, ¹³C, and 2D NMR spectra were recorded using a JEOL ECP 500 MHz (Tokyo, Japan) or an Agilent DD2 500 MHz (Agilent Technologies, Inc., Santa Clara, California, USA) spectrometers.

In vitro antimicrobial assay

Antimicrobial assay for the isolated compounds was performed using an agar well diffusion method towards *E. coli* ATCC 35218, *S. aureus* ATCC 25923, and *C. albicans* ATCC 10231 according to the previous method [30]. The development of microbial growth in a Petri dish was assisted using semisolid media. For bacterial growth, a semisolid medium containing sterilized nutrient agar (NA) (20 g/L) and nutrient broth (NB) (0.8 g/0.1 L) was prepared. Meanwhile, a semisolid medium for fungal growth was prepared with sterilized potato dextrose agar (PDA) (65 g/L) and potato dextrose broth (PDB) (2.4 g/0.1 L). One mL of microbial suspension (turbidity of 0.5 McFarland at 600 nm) was added into NB (5 mL) or PDB (5 mL) and mixed. Solutions of tested compounds were prepared with concentrations of 0.1, 0.2, and 0.3% in DMSO (10%, v/v). Chloramphenicol and ketoconazole each in 0.1% (v/v) were used as respective positive controls for bacteria and fungi, while DMSO was used as a negative control. The solid medium (10 mL) was poured into Petri dishes and allowed to solidify. Onto the solid medium, holes for samples were punched aseptically with tips. After that, the liquid medium-containing microbial suspension was evenly spread onto the surface of the solid medium. Samples (100 μ L) were then added to their respective holes and the Petri dishes were incubated at 37 °C for 24 hours (bacteria) and 72 hours (fungi). Antimicrobial activity was evaluated by measuring the diameter of the clear zone around the hole. The inhibition zone of compounds and positive controls is categorized as strong (10-20 mm), moderate (5-10 mm), and weak (<5 mm).

Results and Discussion

Characterization of isolated compounds

Purification of the methanol extract of the dried powdered *Clathria* sp. using silica gel vacuum liquid chromatography (VLC) and radial chromatography (RC) techniques yielded one new *A*-nor steroid, clathruhoate (1) and other known *A*-nor steroids (2–4). Structures of these compounds are displayed in Figure 2.

RO 4

1 R =
$$COC_3H_7$$
2 R = $COCH_3$
4 R = H

Figure 2. Chemical structures of compounds 1-4

By comparing the values between spectroscopic measurements and literatures data, compounds **2–4** were identified as 3β -(acetoxymethyl)-A-nor- 5α -cholestane (**2**) [25], 3β -(hydroxymethyl)-A-nor- 5α -cholest-15-ene (**3**) [25, 26], and 3β -(hydroxymethyl)-A-nor- 5α -cholestane (**4**) [25]. A-nor sterols are produced by marine Demosponges (e.g Axinella sp., Homoaxinella sp., Phakellia sp.) [25–28] via conversion of dietary sterols [29]. The occurrence of compounds **3** and **4** have been described in species Axinella [27, 28] and Homoaxinella [25], while the acetylation of compound **4** produced compound **2** [25]. Herein we report compound **2** as a natural product of the investigated Clathria sp. To our knowledge, the occurrence of these compounds in the genus Clathria is firstly reported in this work.

3β -(butyryloxymethyl)-*A*-nor- 5α -cholestane (1)

White solid. HRMS-ESI: m/z 459.4112 [M+H]⁺ (calculated for $C_{31}H_{55}O_2^+$: 459.4197). ATR-FTIR (ν_{max} , cm⁻¹): 2922, 2853, 1734, 1464, 1379, 1173, 968, 723. ¹H and ¹³C NMR data are listed in Table 1.

Table 1. NMR data of compounds 1–4 (125 MHz for 13 C and 500 MHz for 1 H, in CDCl₃, δ , ppm, J/Hz)

C atom		1		3	4
C atom	$\delta_{ m C}$	$\delta_{ m H}$	$\delta_{ m C}$	$oldsymbol{\delta}_{ ext{C}}$	$\delta_{ m C}$
1	39.0	1.08 (1H, m), 1.61 (1H, m)	39.1	39.9	39.1
2	27.3	1.44 (1H, m), 1.94 (1H, m)	22.8	22.9	24.6
3	38.5	2.31 (1H, m)	38.6	39.7	42.9
4	68.3	3.94 (1H, dd, J=10.27, 6.85), 4.09 (1H, dd, J=10.27, 6.85)	68.5	67.2	66.7
5	52.4	1.44 (1H, m)	52.6	52.7	52.6
6	24.4	1.04 (1H, m), 1.55 (1H, m)	27.5	27.4	27.4
7	32.7	0.83 (1H, m), 1.74 m (1H, m)	32.8	32.9	32.8
8	35.8	1.36 (1H, m)	35.7	35.7	35.6
9	55.4	0.73 m (1H, m)	55.6	55.6	55.5
10	44.1	-	44.3	44.2	44.1
11	22.8	1.31 (2H, m)	23.4	24.6	22.8
12	39.5	1.14 (2H, m)	39.5	36.3	39.9
13	43.1	-	43.1	43.1	43.1
14	56.3	0.99 (1H, m)	56.4	56.5	56.5
15	23.8	1.14 (1H, m), 1.34 (1H, m)	24.6	130.0	24.6
16	29.3	1.26 (2H, m)	28.3	131.1	28.3
17	56.2	1.00 (1H, m)	56.4	56.4	56.4
18	12.2	0.66 (3H, s)	12.3	12.3	12.1
19	14.5	0.78 (3H, s)	14.6	14.7	14.4
20	35.5	1.29 ((1H, m)	35.9	29.8	35.8
21	18.7	0.91 (3H, m)	18.9	18.9	18.7
22	36.2	0.99 (2H, m)	36.3	35.9	36.2
23	23.2	1.33 (1H, m), 1.44 (1H, m)	23.0	24.6	23.9
24	39.9	1.14 (1H, m), 1.95 (1H, m)	40.0	40.0	39.5
25	28.2	1.24 (1H, m), 1.81 (1H, m)	28.2	28.2	28.0
26	22.7	0.86 (3H, m)	22.7	22.7	22.7
27	22.6	0.86 (3H, m)	22.6	23.0	22.7
1'	174.1	_	171.5		
2'	34.5	2.27 (2H, t, J=7.83)	21.2		
3'	22.5	1.63 (2H, m)			
4'	14.1	0.89 (3H, m)			

Compound 1 was isolated as a white solid and was found to have a molecular formula of $C_{31}H_{55}O_2$ (MW 458.41238) based on a HRESI-MS analysis at m/z 459.4112 [M + H]⁺, requiring five degrees of unsaturation. Of

these, one degree of unsaturation could be attributable to a carbonyl resonance (δ 174.1), while the remaining degrees were denoted by four rings. The IR spectrum showed absorption bands of methyl and methylene groups at 2922 cm⁻¹ (CH₃ stretching), 2853 cm⁻¹ (CH₂ stretching), 1464 cm⁻¹ and 1379 cm⁻¹ (CH₃ and CH₂ deformation) and an ester group at 1734 cm⁻¹ (C=O stretching) and 1173 cm⁻¹ (C=O stretching). The ¹H NMR spectrum showed signals representing five resonances of methyl protons at δ 0.66 (H-18), 0.78 (H-19), 0.86 (H-26, H-27) and 0.91 (H-21) and two double doublet (dd) resonances of oxymethylene protons (H-4) at δ 3.94 (J=10.27, 6.85) and 4.09 (J=10.27, 6.85), indicating a typical 3 β -(hydroxymethyl)-A-nor cholestane [25, 26]. In addition, a methyl group at δ 0.89 (H-4') and two methylenes at δ 1.63 (H-3') and 2.27 (H-2') were observed in the spectrum, suggesting a butyryl group of an ester functionality in the structure as inferred in the IR analysis. In the ¹³C NMR spectrum, thirty-one carbon signals were observed, thus supporting the results from the mass analysis. The carbon of oxymethylene of the A-nor steroid was resonanced at δ 68.3 (C-4) and one carbonyl of ester functionality was resonanced at δ 174.1 (C-1'). The assignments of these protons and carbons in the skeleton of compound 1 were based on a HSQC analysis and are given in Table 1.

The presence of ca holestane skeleton in compound 1 was verified using 2D NMR analyses, in particular, the HMBC measurement. Using this analysis, all carbons with the exception of C-2 (δ 27.3), C-6 (δ 24.4), C-11 (δ 22.8), and C-15 (δ 23.8) could be associated with one or more of the methyl groups based on $^2J_{CH}$ and $^3J_{CH}$ correlations (Figure 3). The patterns of these correlations were similar to the known compounds 2 and 4, which confirmed 1 as an A-nor cholestane. The oxymethylene protons at C-4 and methine proton at C-3 showed $^3J_{CH}$ and $^4J_{CH}$ correlations to a carbonyl carbon (δ 174.1) of an ester functionality, respectively, confirming the position of this carbonyl at C-1'. The presence of a butyryloxy group (C₃H₇COO-) was confirmed using the observed correlations of the methyl (H-4') and methylenes (H-2', H-3') protons toward the carbonyl carbon C-1'. In addition, protons H-2' also showed $^4J_{CH}$ correlation to the oxymethylene carbon (C-4). Hence, this side group was attached to C-4. Based on these evidences, compound 1 was established as 3β -(butyryloxymethyl)-A-nor-5 α -cholestane for which clathruhoate as a trivial name is proposed.

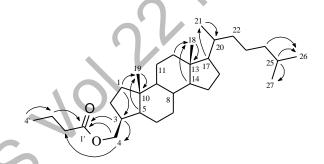


Figure 3. HMBC correlations (H \rightarrow C) of compound 1

3β -(acetoxymethyl)-A-nor-5α-cholestane (2)

Yellow crystal. MW: 430.7061 ($C_{29}H_{50}O_2$). ATR-FTIR (v_{max} , cm⁻¹): 2926, 2858, 1740, 1455, 1370, 1164, 969, 721; ¹H NMR (CDCl₃, 500 MHz, δ/ppm, J/Hz): 4.09 (1H, dd, J=10.27, 7.15, H-4a)), 3.95 (1H, t, J=8.4, H-4b), 2.33 (1H, m, H-3), 2.03 (3H, s, H-2'), 0.91 (3H, m, H-21), 0.86 (3H, m, H-27), 0.85 (3H, m, H-26), 0.75 (3H, s, H-19), 0.65 (3H, s, H-18). ¹³C NMR data are given in Table 1.

Compound **2** was obtained as a yellow crystal and gave a molecular formula of $C_{29}H_{50}O_2$ (MW 430.7061) based on analyses using 1H and ^{13}C NMR spectra. According to this formula, five degrees of unsaturation were calculated as denoted by a carbonyl resonance (δ 171.5) and four rings of the cholestane skeleton. The IR spectrum of compound **2** showed the presence of methyl and methylene groups (2926, 2858, 1455, 1370 cm⁻¹) and an ester group (1740, 1164 cm⁻¹), which was similar to compound **1**. The 1H NMR spectrum exhibited six resonances of methyl protons at δ 0.65 (s, H-18), 0.75 (s, H-19), 0.85 (m, H-26), 0.86 (m, H-27), 0.91 (m, H-21) and 2.03 (s, H-2') and multiplet resonances of typical cholestane protons ranging from δ 0.83 to 2.33. The oxymethylene protons (H-4) were

observed as a double doublet at δ 4.09 (J=10.27, 7.15, H-4a) and a triplet at δ 3.95 (J=8.14, H-4b). Meanwhile, the ¹³C NMR spectra showed twenty-nine carbons in the skeleton of compound **2** (Table 1), and their values were identical to those given in [25]. Hence, compound **2** was deduced as 3β -(acetoxymethyl)-A-nor-5 α -cholestane.

3β -(hydroxymethyl)-A-nor- 5α -cholest-15-ene (3)

White solid. MW: 386.6535 ($C_{27}H_{46}O$). ATR-FTIR (ν_{max} , cm⁻¹): 3334, 2918, 2853, 1594, 1508, 1454, 1376, 1265, 1035, 977. ¹H NMR (CDCl₃, 500 MHz, δ /ppm, J/Hz): 5.15 (1H, m, H-15), 5.13 (1H, m, H-16), 3.71 (1H, dd, J=10.45, 6.50, H-4a), 3.47 (1H, t, J=9.72, H-4b), 2.22 (1H, m, H-3), 1.95 (2H, m), 1.82 (1H, m), 0.90 (3H, m, H-21), 0.89 (3H, m, H-27), 0.85 (3H, m, H-26), 0.74 (3H, s, H-19), 0.65 (3H, s, H-18). ¹³C NMR data are given in Table 1.

Compound **3** was obtained as a white solid. The compound has a molecular formula of $C_{27}H_{46}O$ (MW 386.6535) based on ^{1}H and ^{13}C NMR spectral analyses and required five degrees of unsaturation, originating from one olefinic group (δ 130.0, 131.1) and four rings of the cholestane structure. The IR spectrum displayed absorption bands of a hydroxyl group (3334 cm⁻¹, OH stretching), methyl and methylenes groups (2918, 2853, 1454, 1376 cm⁻¹), and an alkene group (1594 cm⁻¹, C=C stretching). The ^{1}H NMR spectrum of compound **3** exhibited resonances of olefinic protons at δ 5.15 (m, H-15) and 5.13 (m, H-16), oxymethylene protons at δ 3.71 (dd, J=10.45, 6.50, H-4a) and 3.47 (t, J=9.72, H-4b), and methyl protons at δ 0.65 (s, H-18), 0.74 (s, H-19), 0.85 (m, H-26), 0.89 (m, H-27) and 0.90 (m, H-21). The presence of a hydroxyl group in the IR spectrum and oxymethylene protons (δ 3.71, 3.47) in the ^{1}H NMR spectrum suggested an alcohol functionality in compound **3**. The ^{13}C NMR spectrum consisted of twenty-seven carbons including two olefinic carbons (δ 130.0, 131.1) and one oxymethylene carbon (δ 67.2) (Table 1). The NMR data of compound **3** were identical with those assigned for 3 β -(hydroxymethyl)-A-nor-5 α -cholest-15-ene [25, 26]. This compound was previously reported from *Homoaxinella trachys* [25], *Phakellia aruensis* [26], and *Axinella proliferans* [27].

3β -(hydroxymethyl)-*A*-nor- 5α -cholestane (4)

White solid. MW: 388.6694 ($C_{27}H_{48}O_2$). ATR-IR (ν_{max} , cm⁻¹): 3357, 2915, 2820, 1454, 1378, 1276, 1196, 740; ¹H NMR (CDCl₃, 500 MHz, δ /ppm, J/Hz): 3.70 (1H, dd, J=10.3, 6.3, H-4a), 3.46 (1H, t, J=9.6, H-4b), 2.23 (1H, m, H-3), 1.93 (2H, m), 1.82 (1H, m), 0.91 (3H, m, H-21), 0.86 (3H, m, H-27), 0.84 (3H, m, H-26), 0.73 (3H, s, H-19), 0.64 (3H, s, H-18). ¹³C NMR data are given in Table 1.

Compound 4 was isolated as a white solid and have a molecular formula of $C_{27}H_{48}O_2$ (MW 388.6694). Four degrees of unsaturation calculated from this formula was attributed by four rings of the cholestane structure. The IR spectrum suggested a hydroxyl group (3357 cm⁻¹) and methyl and methylenes groups (2918, 2853, 1454, 1376 cm⁻¹). The ¹H NMR spectrum showed two coupled peaks at δ 3.70 (dd, J=10.3, 6.3) and 3.46 (t, J=9.6), indicating the mutual oxymethylene protons as previously observed in compounds 1–3. Similarly, five methyl groups of typical cholestane skeleton were observed at δ 0.64 (s, H-18), 0.73 (s, H-19), 0.84 (m, H-26), 0.86 (m, H-27) and 0.91 (m, H-21). Meanwhile, as many as twenty-seven carbons were obtained from the ¹³C NMR spectrum and their values were identical to compounds 1–2 with the exception of butyryloxy and acetoxy carbons. Hence, compound 4 was determined as 3β -(hydroxymethyl)-A-nor- 5α -cholestane [25]. This compound was previously reported from *Axinella verrucosa* [28].

Antimicrobial activity of isolated compounds

The isolated compounds were evaluated for antibacterial activity against *E. coli*, *S. aureus*, and antifungal activity against *C. albicans*. However, none of the tested compounds exhibited antibacterial and antifungal activities as compared to chloramphenical and ketoconazole, respectively. To our knowledge, the antimicrobial activity of these compounds is reported for the first time in this study.

Conclusion

Four A-nor steroids were isolated and identified from the methanol extract of the marine sponge *Clathria* sp. in the genus *Clathria* for the first time, which are 3β -(butyryloxymethyl)-A-nor- 5α -cholestane (1), 3β -(acetoxymethyl)-A-nor- 5α -cholestane (2), 3β -(hydroxymethyl)-A-nor- 5α -cholestane (3), and 3β -(hydroxymethyl)-A-nor- 5α -cholestane (4). Compound 1 named as clathruoate was established as a new compound from this species. The

presence of these compounds could enrich the chemical diversity of the genus *Clathria*. Despite their inactive properties towards *E. coli*, *S. aureus*, and *C. albicans*, further studies are still required to explore more about the potent antimicrobial compounds from this species as well as other biological activities.

Acknowledgment

The authors thank the Ministry of Research, Technology, and Higher Education of the Republic of Indonesia for a research grant scheme (Hibah Kompetensi 2016) for financial support.

References

- 1. Spalding, M. D., Ravilious, C. and Green, E. P. (2001). World atlas of coral reefs. University of California Press, Berkeley, Los Angeles, CA: pp. 21 22.
- 2. Mora, C., Chittaro, P. M., Sale, P. F., Kritzer, J.P. and Ludsin, S.A. (2003). Patterns and processes in reef fish diversity. *Nature*, 421: 933 936.
- 3. van Soest, R. W. M. (1989). The Indonesia sponge fauna: a status report. *Netherlands Journal of Sea Research*, 23: 223 230.
- 4. Soekarno, R. (1989). Comparative studies on the status of Indonesia coral reefs. *Netherlands Journal of Sea Research*, 23: 215 222.
- 5. van Soest, R. (2017). *Clathria* Schmidt, 1982. In *Wold Porifera Database*. van Soest, R. W. M., Boury-Esnault, N., Hooper, J. N. A., Rützler, K., de Voogd, N. J., Alvarez de Glasby, B., Pisera, A. B., Manconi, R., Schoenberg, C., Klautau, M., Picton, B., Kelly, M., Vacelet, J., Dohrmann, M., Díaz, M. C., Cárdenas, P., Carballo, J. L. and Rios Lopez, P. (Eds). Accessed through: World Register of Marine Species (http://www.marinespecies.org/aphia.php?p=taxdetails&id=131868 on 2017-07-23).
- 6. Sperry, S. and Crews, P. (1996). A novel alkaloid from the Indo-Pacific sponge *Clathria basilana*. *Tetrahedron Letters*, 37: 2389 2390.
- 7. Capon, R. J., Miller, M. and Rooney, F. (2001). Mirabilin G: a new alkaloid from a southern Australian marine sponge, *Clathria* species. *Journal of Natural Products*, 64: 643 644.
- 8. Zuleta, I. A., Vitelli, M. L., Baggio, R., Garland, M. T., Seldes, A. M. and Palermo, J. A. (2002). Novel pteridine alkaloids from the sponge *Clathria* sp. *Tetrahedron*, 58: 4481 4486.
- 9. El-Naggar, M., Conte, M. and Capon, R. J. (2010). Mirabilins revisited: polyketide alkaloids from a southern Australian marine sponge, *Clathria* sp. *Organic & Biomolecular Chemistry*, 8: 407 412.
- 10. Laville, R., Thomas, O. P., Berrué, F., Marquez, D., Vacelet, J. and Amade, P. (2009). Bioactive guanidine alkaloids from two Carribean marine sponges. *Journal of Natural Products*, 72: 1589 1594.
- 11. Sun, X., Sun, S., Ference, C., Zhu, W., Zhou, N., Zhang, Y. and Zhou, K. (2015). A potent antimicrobial compound isolated from *Clathria cervicornis*. *Bioorganic & Medicinal Chemistry Letters*, 25: 67 69.
- 12. Wei, X., Henriksen, N. M., Skalicky, J. J., Harper, M. K., Cheatham, T. E., III, Ireland, C. M. and Van Wagoner, R. M. (2011). Araiosamines A-D: tris-bromoindole cyclic guanidine alkaloids from the marine sponge *Clathria (Thalysias) araiosa*. *The Journal of Organic Chemistry*, 76: 5515 5523.
- 13. Ohta, S., Okada, H., Kobayashi, H., Oclarit, J. M. and Ikegami, S. (1993). Clathrynamides A, B, and C: novel amides from a marine sponge *Clathria* sp. that inhibit cell division of fertilized starfish eggs. *Tetrahedron Letters*, 34: 5935 5938.
- 14. Davis, R. A., Mangalindan, G. C., Bojo, Z. P., Antemano, R. R., Rodriguez, N. O., Concepcion, G. P., Samson, S. C., de Guzman, D., Cruz, L. J., Tasdemir, D., Harper, M. K., Feng, X., Carter, G. T. and Ireland, C. M. (2004). Microcionamides A and B, bioactive peptides from the Philippine sponge *Clathria (Thalysias) abietina*. *The Journal of Organic Chemistry*, 69: 4170 4176.
- 15. Woo, J. –K., Jeon, J., Kim, C. –K., Sim, C. J., Oh, D. –C., Oh, K. –B. and Shin, J. (2013). Gombamide A, a cyclic thiopeptide from the sponge *Clathria gombawuiensis*. *Journal of Natural Products*, 76: 1380 1383.
- 16. Khokhar, S., Pierens, G. K., Hooper, J. N. A., Ekins, M. G., Feng, Y. and Davis, R. A. (2016). Rhodocomatulin-type anthraquinones from the Australian marine invertebrates *Clathria hirsuta* and *Comatula rotalaria*. *Journal of Natural Products*, 79: 946 953.
- 17. Capon, R. J., Miller, M. and Rooney, F. (2000). Clathrins A-C: metabolites from a southern Australian marine sponge, *Clathria* species. *Journal of Natural Products*, 63: 821 824.
- 18. Rudi, A., Yosief, T., Loya, S., Hizi, A., Schleyer, M. and Kashman, Y. (2001). Clathsterol, a novel anti-HIV-1 RT sulfated sterol from the sponge *Clathria* species. *Journal of Natural Products*, 64: 1451 1453.

- 19. Keyzers, R. A., Northcote, P. T. and Webb, V. (2002). Clathriol, a novel polyoxygenated 14β steroid isolated from the New Zealand marine sponge *Clathria lissosclera*. *Journal of Natural Products*, 65: 598 600.
- 20. Santalova, E. A., Makarieva, T. N., Gorshkova, I. A., Dmitrenok, A. S., Krasokhin, V. B. and Stonik, V. A. (2004). Sterols from six marine sponges. *Biochemical Systematics and Ecology*, 32: 153 167.
- 21. Gupta, P., Sharma, U., Schulz, T. C., McLean, A. B., Robins, A. J. and West, L. M. (2012). Bicyclic C21 terpenoids from the marine sponge *Clathria compressa*. *Journal of Natural Products*, 75: 1223 1227.
- 22. Woo, J. –K., Kim, C. –K., Kim, S. –H., Kim, H., Oh, D. –C., Oh, K. –B. and Shin, J. (2014). Gombaspiroketals A-C, sesterterpenes from the sponge *Clathria gombawuiensis*. *Organic Letters*, 16: 2826 2829.
- 23. Woo, J. –K., Kim, C. –K., Ahn, C. –H., Oh, D. –C., Oh, K. –B. and Shin, J. (2015). Additional sesterterpenes and a nortriterpene saponin from the sponge *Clathria gombawuiensis*. *Journal of Natural Products*, 78: 218 224
- Inbaneson, S. J. and Ravikumar, S. (2012). In vitro antiplasmodial activity of Clathria vulpina sponge associated bacteria against Plasmodium falciparum. Asian Pacific Journal of Tropical Disease, 2012: 319 – 323
- 25. Eggersdorfer, M. L., Kokke, W. C. M. C., Crandell, C. W., Hochlowski, J. E. and Djerassi, C. (1982). Sterols in marine invertebrates. 32. Isolation of 3β -(hydroxymethyl)-A-nor- 5α -cholest-15-ene, the first naturally occurring sterol with a 15-16 double bond. *The Journal of Organic Chemistry*, 47: 5304 5309.
- 26. Malik, S. and Djerassi, C. (1989). Minor and trace sterols in marine invertebrates. 61. Isolation and structure elucidation of new A-nor sterols from the marine sponge *Phakellia aruensis*. Steroids, 53: 271 284.
- 27. Aknin, M., Gaydou, E. M., Boury-Esnault, N., Constantino, V., Fattorusso, E. and Mangoni, A. (1996). Norsterols in *Axinella proliferans*, sponge from the Indian Ocean. *Comparative Biochemistry and Physiology*, 113: 845 848.
- 28. Minale, L. and Sodano, G. (1974). Marine sterols: unique 3β -hydroxymethyl-A-nor- 5α -steranes from the sponge *Axinella verrucosa*. *Journal of the Chemical Society, Perkin Transactions I*: 2380 2384.
- 29. Bibolini, L., Minale, L. and Sodano, G. (1978). Investigations on the ring contraction step in the biosynthesis of A-nor-stanols by the marine sponge *Axinella verrucosa*. *Journal of the Chemical Society, Chemistry Communications*: 524 525.
- 30. Ngajow, M., Abidjulua, J. and Kamu, VS. (2013). Pengaruh antibakteri ekstrak kulit batang Matoa (*Pometia pinnata*) terhadap bakteri *Staphylococcus aureus* secara *in vitro*. *Jurnal Mipa Unsrat Online*, 2(2): 128 132.