

ISOLATION OF CHEMICAL CONSTITUENTS FROM RHIZOMES OF ETLINGERA SPHAEROCEPHALA VAR. GRANDIFLORA

(Pemencilan Kandungan Kimia Daripada Rizom Etlingera Sphaerocephala var. Grandiflora)

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Abstract

A phytochemical investigation was conducted on the rhizomes of *Etlingera sphaerocephala* var. *grandiflora* (Zingiberaceae). Steroids of β -sitosterol and stigmasterol, together with a phenolic compound of paeonol were isolated using several chromatographic means. Their structures were established on the basis of extensive spectroscopic (IR, MS, NMR) data analysis and by comparison of the data obtained with those of the literature.

Keywords: Zingiberaceae, *Etlingera sphaerocephala* var. *grandiflora*, β-sitosterol, stigmasterol, paeonol, NMR

Abstrak

Suatu penyelidikan fitokimia dilakukan ke atas rizom *Etlingera sphaerocephala* var. *grandiflora* (Zingiberaceae). Steroid β-sitosterol dan stigmasterol, berserta sebatian fenolik paeonol dipisahkan menggunakan beberapa cara kromatografi. Struktur mereka dibuktikan berasas kepada analisis data spektroskopi (IM, SJ, RMN) yang ekstensif dan secara membandingkan data tersebut dengan data kepustakaan

Kata kunci: Zingiberaceae, Etlingera sphaerocephala var. grandiflora, β-sitosterol, stigmasterol, paeonol, NMR

Introduction

There are 151 species of Zingiberaceae belonging to 18 genera found in Peninsular Malaysia [1]. The largest genus of Zingiberaceae is *Alpinia* (23 species), followed among others by *Scaphochlamys* (20), *Amomum* (18), *Globba* (15), *Geostachys* (13), *Etlingera* (10) and *Zingiber* (10). The species of *Etlingera* in Peninsular Malaysia has now increased to fifteen [2]. The largest among *Etlingera* species are about 10 m in height. There are more than 100 species of Indo-pacific *Etlingera* growing from sea level to 2500 m [3].

Etlingera sphaerocephala var. grandiflora was described by Holttum [4] as a plant with subterranean, sub-globose, sub-cylindric inflorescence where its flowers appearing at soil level; its stature is 2.5 m; its leaves when young are also suffused red or purple below; its labellum of deep crimson lip, white margins, is 6 cm or more in length and 2.7 cm wide; and the base is red in color. Etlingera sphaerocephala var. grandiflora can be found in many parts of the Peninsular Malaysia and Borneo [5] mainly in lowland forests and at moderate elevation on the mountains. No use has ever been recorded for E. sphaerocephala var. grandiflora [3]. In this paper, we report the isolation and characterization of β-sitosterol, stigmasterol and paeonol from the rhizomes of E. sphaerocephala var. grandiflora. These compounds are reported for the first time from Etlingera.

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Materials and Methods

Plant Material

Rhizomes of the *Etlingera sphaerocephala* var. *grandiflora* were collected in January 2009 from Genting Peras, Hulu Langat, Selangor, Malaysia. A voucher specimen of WYA 386 for the plant has been deposited at the Universiti Kebangsaan Malaysia Herbarium.

Extraction and Isolation

Air-dried rhizomes (315 g) were ground into a fine powder and soaked in n-hexane (5.0 L) for six days. The solution was filtered, and the combined filtrates were rotary evaporated under reduced pressure to produce 1.5 g (0.48%) of a brown-gummy rhizomes n-hexane extract. The remaining rhizomes powder was soaked in methanol (2.5 L) for another six days. The solution was filtered, and the combined filtrates were concentrated using a rotary evaporator to yield 6.3 g (2.1%) of a brown-oily rhizomes successive methanol extract.

The rhizomes n-hexane extract (1.5 g) was subjected to silica gel column chromatography (CC) (Merck 1.09385) by eluting n-hexane containing increasing percentages of ethyl acetate and the fractions collected was 100 mL each. Fractions 4 and 5 were combined (96 mg) and then subjected to radial chromatography (RC) (Merck 1.07749) by using increasing polarities of n-hexane: ethyl acetate to produce 12.4 mg of β -sitosterol (1). Fractions 6, 7 and 8 were combined (139 mg) and then subjected to silica gel preparative thin layer chromatography (PTLC) (Merck 1.07749) using 100:0 to 10:90 of n-hexane: ethyl acetate to afford 6.6 mg of stigmasterol (2).

The rhizomes successive methanol extract (6.3 g) underwent silica gel flash CC (Merck 1.07747) using 10% polarity increments from 90: 10 chloroform: methanol to 100% methanol whereby 100 mL fractions were collected. The flash CC fractions 5, 6 and 7 were combined (170 mg) and subjected to RC to produce five fractions. The combined fractions 1, 2 and 3 were fractionated over silica gel (Merck 1.07749) on PTLC to give 2.4 mg of paeonol (3).

β-Sitosterol (1; si) /Stigmasterol (2; st)

Colorless needles (12.4 mg)/white powder (6.6 mg). IR cm⁻¹: 3393/3434, 2930/2924, 1611/1639, 1457/1457, 1385/1375, 1171/1179. EIMS for β-sitosterol-C₂₉H₅₀O m/z (rel. int.): 414 [M⁺] (100%), 396 (30.8%), 381 (14.0%), 329 (13.7%), 303 (22.1%), 255 (11.8%), 213 (12.2%), 145 (18.5%), 95 (21.6%), 81 (21.3%), 55 (25.8%), 43 (45.0%). EIMS for stigmasterol-C₂₉H₄₈O m/z (rel. int.): 412 [M⁺] (39.7%), 351 (13.5%), 314 (7.0%), 300 (25.5%), 271 (38.4%), 229 (8.6%), 213 (10.6%), 55 (100%). ¹H NMR (CDCl₃, 400 MHz): 3.53 (1H, m, H-3), 5.36 (1H, d, J = 5.1/7.3 Hz, H-6), 0.69 (3H, s, H-18), 1.02 (3H, s, H-19), 0.93 (3H, d, J = 6.6/6.2 Hz, H-21), 5.16_{st} (1H, dd, J = 15.6, 8.4 Hz, H-22); 5.02_{st} (1H, dd, J = 15.6, 9.2 Hz, H-23), 0.85 (3H, d, J = 7.0/7.3 Hz, H-26), 0.83/0.81 (3H, d, J = 7.0 Hz, H-27), 0.87/0.83 (3H, t, J = 7.7/7.0 Hz, H-29), 1.05-2.32 (others). ¹³C NMR (CDCl₃, 100 MHz): 141.0 (C-5), 121.9 (C-6), 72.0 (C-3), 57.0/57.1 (C-14), 56.3 (C-17), 50.4/50.3 (C-9), 46.1/51.5 (C-24), 42.6/42.3 (C-4, C-13), 40.0 (C-12), 37.5 (C-1), 36.7 (C-10), 36.4/40.7 (C-20), 34.2/138.5 (C-22), 32.2/32.1 (C-8), 32.1_{si} (C-7), 31.9_{st} (C-7, C-25), 31.9/31.7 (C-2), 29.4_{si} (C-25), 28.5/29.3 (C-16), 26.3/129.5 (C-23), 24.5 (C-15), 23.3/25.6 (C-28), 21.3_{si} (C-11), 21.3_{st} (C-11, C-21), 20.1/19.0 (C-27), 19.6 (C-19), 19.3/21.4 (C-26), 19.0_{si} (C-21), 12.2 (C-29), 12.1 (C-18).

Paeonol (3)

White needles (2.4 mg). IR cm $^{-1}$: 3432, 2925, 1720, 1383, 1246, 1021. EIMS for C₉H₁₀O₃ m/z (rel. int.): 166 [M $^{+}$] (31.3%), 151 (100%), 108 (16.5%), 95 (21.0%), 43 (34.6%). H NMR (400 MHz, CDCl₃) δ : 2.56 (3H, s, CH₃), 3.84 (3H, s, OCH₃), 6.42-6.45 (2H, m, H-3, H-5), 7.63 (1H, d, J=8.8 Hz, H-6), 12.75 (1H, s, OH). CDCl₃) δ : 114.1 (C-1), 165.4 (C-2), 101.0 (C-3), 166.3 (C-4), 107.9 (C-5), 132.5 (C-6), 202.8 (C=O), 55.8 (OCH₃), 26.4 (CH₃).

2 (trans-CH=CH across C22--C23; Stigmasterol)

Results and Discussion

3 (Paeonol)

Purification of the Etlingera sphaerocephala var. grandiflora rhizomes extracts produced β-sitosterol (1), stigmasterol (2) and paeonol (3). β-Sitosterol (1) was isolated as colorless needles. Column chromatography of the rhizomes n-hexane extract gave a 139 mg combined fractions, and its preparative thin layer chromatography yielded 6.6 mg of stigmasterol (2) as white powder. Spectral data of stigmasterol differed minimally from those of βsitosterol at and around a trans-C22-C23 double bond in the former. The EIMS of β-sitosterol/stigmasterol showed the molecular ion m/z 414/412 [M⁺] that corresponds to the formula C₂₉H₅₀O/C₂₉H₄₈O and in agreement with other previous spectral data. H NMR spectrum of β-sitosterol/stigmasterol indicated the presence of six methyl peaks of H-18, H-27, H-29, H-26, H-21 and H-19 that appeared at respective δ 0.69, 0.83/0.81, 0.87/0.83, 0.85, 0.93 and 1.02. The hydroxymethine proton H-3 of β -sitosterol/stigmasterol came out at δ 3.53 as a multiplet. One proton appeared at δ 5.36 as a doublet with coupling constants of 5.1/7.3 Hz represents the endocyclic double bond proton H-6 of β-sitosterol/stigmasterol. Stigmasterol showed two doublet-of-doublets for the other olefinic protons H-23 and H-22 at δ 5.02 (1H, dd, J = 9.2, 15.6 Hz) and 5.16 (1H, dd, J = 8.4, 15.6 Hz) respectively. The ¹H NMR spectral data of stigmasterol are in agreement with those of Hussain et al. [6]. The ¹³C NMR spectrum of βsitosterol/stigmasterol showed 28/26 signals for the 29 carbons skeleton with the overlapping of 2/6 carbons. The significant carbon signal for the β-sitosterol/stigmasterol would be the C-3 attached to a hydroxyl group and appeared at δ 72.0. The endocyclic carbon-carbon double bond of β-sitosterol/stigmasterol was represented by two signals at δ 121.9 and 141.0 of C-6 and C-5. Other olefinic carbons of stigmasterol appeared at δ 138.5 and 129.5 for C-22 and C-23. The quaternary C-5 signal is shifted to lower field than those of the tertiary C-6, C-22 and C-23. The similarity of the ¹³C spectral data of β-sitosterol/stigmasterol with those of the published data in [7]/[8]

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confirmed that both two are of the same structure. β-Sitosterol/stigmasterol has never been reported before from any other *Etlingera*. However β-sitosterol was found in many other plant families including Zingiberaceae such as *Zingiber* [9], *Alpinia* [10], *Globba* [11], *Kaempferia* [12], *Costus* [13], *Renealmia* [14], *Aframomum* [15] and *Hedychium* [16], whereas stigmasterol including Zingiberaceae of *Renealmia* [14], *Alpinia* [17, 18] and *Zingiber* [9].

Paeonol (3) was isolated from the rhizomes successive methanol extract as white needles. Mass spectrum of this compound showed a molecular ion peak at m/z 166, which is in agreement with the molecular formula C₉H₁₀O₃ of paeonol. ¹H NMR spectrum of paeonol showed that there are two singlets at δ 2.56 and 3.84 representing two methyl groups attached to respective quaternary carbonyl, and oxygen. A signal for H-3, H-5 appeared at δ 6.42-6.45 as a multiplet. Proton H-6 showed a doublet which appeared at δ 7.63 with a coupling constant of 8.8 Hz. A hydrogen-bonded, six-membered ring phenolic proton was displayed very down-field at δ 12.75. ¹³C NMR data of this compound showed that there were nine signals which represents nine different carbons. The signal at δ 202.8 was assigned to the carbonyl group of C-1. Moreover, signals at δ 101.0, 107.9 and 132.5 were assigned to respective aromatic carbons of C-3, C-5 and C-6. Signal at δ 165.4 was assigned to C-2 which is known as an aromatic carbon attached to a hydroxyl group whereas the appearance of the aromatic C-4 with the methoxy substitution was seen at δ 166.3 due to the electron donating feature of the oxygen atoms in those groups. Two upfield signals appeared at δ 26.4 and 55.8 for methyl groups of COCH₃ and OCH₃. Comparison of the ¹H and ¹³C NMR data of paeonol with the previously published data in [19] confirmed that both are of the same compound. Paeonol has never been isolated before from any genus of Zingiberaceae. The fact that paeonol was found in the woody plants of Luculia intermedia [20], Rosmarinus officinalis [21], Moutan cortex [22] as well as in the nonwoody plants of Arisaema erubescens [23], Exacum affine [24], Primula auricula [25], Cynanchum paniculatum [25] supports its existence in our non-woody plant of Etlingera sphaerocephala var. grandiflora.

Conclusion

The isolation and identification of β -sitosterol (1), stigmasterol (2) and paeonol (3) from the rhizomes of *Etlingera* sphaerocephala var. grandiflora was the first ever to be reported from this plant. The work was carried out by utilizing several kinds of chromatographic separation techniques and spectroscopic analyses.

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References

- 1. Turner, I.M. (1995). A catalogue of the vascular plants of Malaya. *Gardens' Bulletin Singapore*. 47(2): 642-655.
- 2. Lim, C.K. (2001). Taxonomic notes on *Etlingera* Giseke (Zingiberaceae) in Peninsular Malaysia: the "*Achasma*" taxa and supplementary notes on the "*Nicolaia*" taxa. *Folia Malaysiana*. 2(3): 141-178, 209-210.
- 3. Poulsen, A.D. (2006). Etlingera of Borneo. Natural History Publications (Borneo). Kota Kinabalu.
- 4. Holttum, R.E. (1950). The Zingiberaceae of the Malay Peninsula. *Gardens' Bulletin Singapore*. 13(1): 1-249.
- 5. Smith, R.M. (1986). New combinations in *Etlingera* Giseke (Zingiberaceae). *Notes Royal Botanic Garden Edinburgh*. 43: 243-254.
- 6. Hussain, M.M., Rahman, M.S., Jabbar, A., Rashid, M.A. (2008). Phytochemical and biological investigations of *Albizzia lebbeck* Benth. *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas*. 7(5): 273-278.
- Zhang, X., Geoffroy, P., Miesch, M., Julien-David, D., Raul, F., Aoude-Werner, D., Marchioni, E. (2005). Gram-scale chromatographic purification of β-sitosterol. Synthesis and characterization of β-sitosterol oxides. Steroids. 70(13): 886-895.
- 8. De-Eknamkul, W., Potduang, B. (2003). Biosynthesis of β-sitosterol and stigmasterol in *Croton sublyratus* proceeds via a mixed origin of isoprene units. *Phytochemistry*. 62: 389-398.

- 9. Peng, W., Zhang, Y., Yang, K., Xiao, Y. (2007). Chemical constituents of *Zingiber officinale* (Zingiberaceae). *Yunnan Zhiwu Yanjiu*. 29(1): 125-128.
- 10. Phan, M.G., Le, H.T., Phan, T.S., Otsuka, H. (2007). Chemical constituents of the fruits of *Alpinia conchigera* Griff. (Zingiberaceae). *Tap Chi Hoa Hoc*. 45(4): 509-512.
- 11. Qiao, C., Xu, L., Shen, Y., Hao, X. (1999). Chemical constituents from *Globba racemosa* Smith. *Huaxue Yanjiu Yu Yingyong*. 11(5): 575-576.
- 12. Sukari, M.A., Neoh, B.K., Lajis, N.H., Ee, G.C.L., Rahmani, M., Ahmad, F.B.H., Yusof, U.K. (2004). Chemical constituents of *Kaempferia angustifolia* (Zingiberaceae). *Oriental Journal of Chemistry*. 20(3): 451-456.
- 13. Qiao, C., Li, Q., Dong, H., Xu, L., Wang, Z. (2002). Studies on chemical constituents of two plants from *Costus. Zhongguo Zhongyao Zazhi*. 27(2): 123-125.
- 14. Lognay, G., Marlier, M., Haubruge, E., Trevejo, E., Severin, M. (1989). Study of the lipids from *Renealmia alpinia* (Rott) Maas. *Grasas y Aceites*. 40(6): 351-355.
- 15. Vidari, G., Vita Finzi, P., De Bernardi, M. (1971). Flavonols and quinones in stems of *Aframomum giganteum*. *Phytochemistry*. 10(12): 3335-3339.
- 16. Xiao, P., Sun, C., Zahid, M., Ishrud, O., Pan, Y. (2001). New diterpene from *Hedychium villosum*. *Fitoterapia*. 72(7): 837-838.
- 17. Phan, M.G., Phan, T.S. (2004). Phytochemical investigation of *Alpinia globosa* (Lour.) Horaninov, Zingiberaceae. *Tap Chi Hoa Hoc*. 42(3): 376-378.
- 18. Phan, M.G., Dang, B.T., Phan, T.S. (2005). Flavonoid compounds from the rhizomes of *Alpinia conchigera* Griff., Zingiberaceae. *Tap Chi Hoa Hoc*. 43(1): 105-108.
- 19. Kim, M.Y., Lee, H.S., Bae, K.H., Yun, Y.S., Song, J.Y., Han, Y.S. (2006). Use of paeonol for inhibiting angiogenesis or for enhancing radiosensitization. PCT Int. Appl. WO 2006121263 A1 20061116.
- 20. Zhou, F., Tan, G., Liang, P. (1981). Isolation of paeonol, an analgesic principle from *Luculia intermedia* Hutch. *Zhongcaoyao*. 12(7): 6.
- 21. Cheng, W., Chen, H., Zhang, Y., Gao, S., Tao, Y., Gu, K. (2005). Study on chemical constituents in *Rosmarinus officinalis. Zhongcaoyao*. 36(11): 1622-1624.
- 22. Kitagawa, I., Yoshikawa, M., Tsunaga, K., Tani, T. (1979). Studies on *Moutan cortex*. II. On the chemical constituents. *Shoyakugaku Zasshi*. 33(3): 171-177.
- 23. Ducki, S., Hadfield, J.A., Lawrence, N.J., Zang, X., McGown, A.T. (1995). Isolation of paeonol from *Arisaema erubescens*. *Planta Medica*. 61(6): 586-587.
- 24. Matsumoto, M. (1994). 2'-Hydroxy-4'-methoxyacetophenone (paeonol) in *Exacum affine* cv. *Bioscience*, *Biotechnology*, *and Biochemistry*. 58(10): 1892-1893.
- 25. Weeks, R.A., Dobberstein, R.H., Farnsworth, N.R. (1977). Isolation of paeonol from *Bathysa meridionalis*. *Lloydia*. 40(5): 515-516.